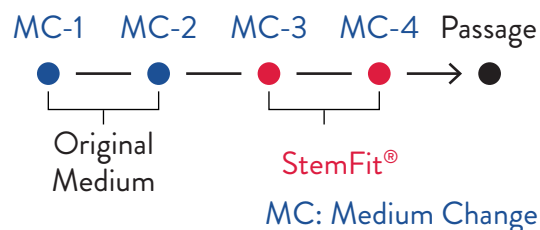


TIPS ON TRANSITIONING CELLS TO STEMFIT® MEDIUM

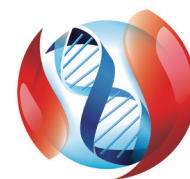
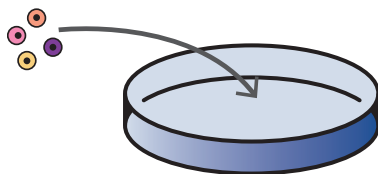
FEEDER FREE MEDIUM FOR ES/IPS CELLS

Key points for successful single-cell passage

- Switch culture medium to StemFit® 2-3 days prior to passage



- Seed the cells at higher density ($>1.0 \times 10^5$ cells per well (6-well plate))



StemFit® Technical Tips

**ROBUST AND
REPRODUCIBLE CULTURE**

Quantitative culture

HIGH FOLD EXPANSION

~100X expansion / passage

AMSBIO | www.amsbio.com | info@amsbio.com

AMSBIO LLC
USA & Canada

1035 Cambridge Street,
Cambridge, MA 02141
T: +1 (617) 945-5033 or
T: +1 (800) 987-0985
F: +1 (617) 945-8218

AMSBIO Europe BV
EU

Berenkoog 41,
1822 BH Alkmaar,
Netherlands
T: +31 (0) 72 8080244
F: +31 (0) 72 8080142

AMS Biotechnology (Europe) Ltd
UK & Rest of the World

184 Park Drive, Milton Park
Abingdon OX14 4SE
T: +44 (0) 1235 828 200
F: +44 (0) 1235 820 482

AMS Biotechnology (Europe) Ltd
Switzerland

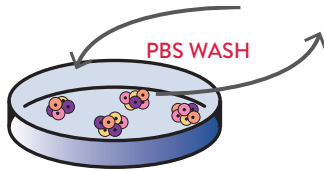
Via Lisano 3,
(CP.683)
CH-6900 Massagno
T: +41 (0) 91 604 55 22
F: +41 (0) 91 605 17 85

AMSBIO Europe BV
Deutschland

T: +49 (0) 69 779099
F: +49 (0) 69 13376880

SINGLE-CELL PASSAGE BRIEF PROTOCOL EXAMPLE (6-WELL PLATE) AND TIPS

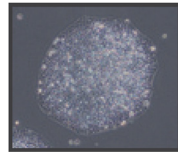
1. Aspirate the medium and wash once with 2ml of PBS



2. Add 500 μ l/well of Accutase and incubate at 37°C for 10 minutes

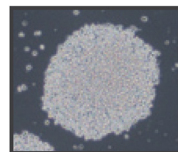
- TrypLE™ can also be used for cell dissociation
- Incubation time may vary depending on the matrix

- Before incubation with Accutase



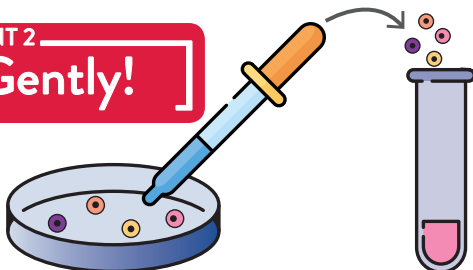
POINT 1
[10 min!]

- Gaps in the colonies appear and dissociation of colonies is apparent



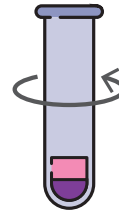
3. Gently pipette the cells to fully dissociate and transfer cells to a 15ml tube filled with 500 μ l of culture medium containing 10 μ M Y-27632

POINT 2
[Gently!]



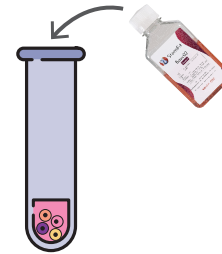
4. Count the cells and centrifuge the tubes

[300g RT 4 min]



Aspirate the medium and resuspend cells with culture medium containing 10 μ M Y-27632

[1000 cells / μ l]

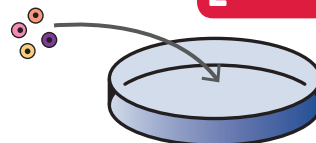


5. Add 10-20 μ l ($1.0-2.0 \times 10^4$ cells) of resuspended cells per well in 1.5 mL of culture medium containing 10 μ M Y-27632

- It is important to adjust the plating cell number for different lines of hPSCs
- Try higher seeding density when cell or colony quantity is insufficient

POINT 3
[Adjust the cell number]

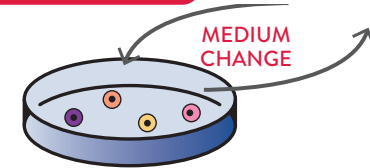
POINT 3
[Distribute evenly!]



- Immediately distribute the cells evenly over the plate surface to avoid uneven attachment

6. After >24 hours of culture, replace with fresh culture medium without Y-27632

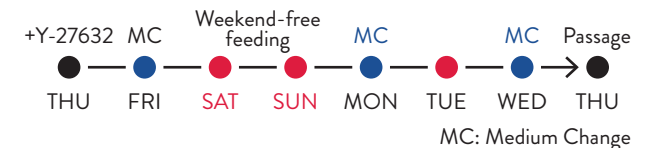
POINT 5
[>24 hours!]



- It is critical that cells are cultured in Y-27632 containing medium for more than 24 hours

7. Perform medium change

< Passage Schedule Example >



POINT 6
[* If the colour of the medium turns orange or yellow it should be changed every day]

POINT 7
[* Do not allow cells to become confluent]