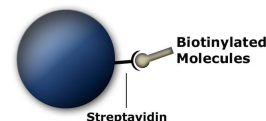




MagSi-STA 600, 1.0

Product Description



I. Intended use

MagSi-STA beads are useful for many applications, including purification of proteins and peptides or nucleic acids, immunoprecipitation, immunoassays, protein interaction studies, phage display, and cell isolation. MagSi-STA beads are coated with streptavidin, which efficiently binds to biotinylated molecules, e.g. peptides, proteins, antibodies, sugars, lectins. The magnetic properties enable easy and quick washing steps. Because beads are in suspension, incubation times can be shortened compared to alternative techniques. Columns or centrifugation steps are not necessary when working with magnetic beads. This feature enables easy implementation into automated processes.

Binding of molecules onto MagSi-STA is based on the strong non-covalent interaction between streptavidin and biotin ($K_a = 10^{15} \text{ M}^{-1}$). Biotin is easily conjugated to various molecules and inexpensive biotinylated products are sold by many companies. MagSi-STA beads are coated with recombinant streptavidin (53 kDa) with shortened N- and C-terminus for improved solubility and accessibility of the sites. Albumin binding sites are eliminated for optimal specificity.

MagSi-STA beads are offered in sizes of 600 nm and 1.0 μm . The sedimentation time of 600nm MagSi beads has been optimized and is approx. 4 times compared to 1.0 μm beads. This allows e.g. long incubation times without shaking/mixing etc. MagSi beads with a diameter of 1 μm have stronger magnetic properties and will separate approx. 2x faster compared to 600nm in the same conditions; the typical separation time is ≤ 1 minute using a suitable magnet.

II. Principle

MagSi-STA beads are added to a sample containing biotinylated molecules. The biotinylated molecules will bind to the beads during a short incubation. The complex is separated from the sample using a magnet and can be used in downstream applications.

III. Material Supplied

- 2, 10, or 100 ml MagSi-STA 600 or MagSi-STA 1.0 (supplied at 10 mg/ml in PBS, 0.05% Tween20, 0.05% sodium azide)

Additional materials needed:

- Buffers and Materials (depending on the application, contact for support)
- Magnetic separator for bead separation/collecting (see order information)
- Mixer/vortex to homogenize samples and resuspend beads (depending on the application, contact for support)

Washing and Binding Buffers :

- For coupling of proteins and peptides a neutral buffer (PBS) is recommended, optionally with a surfactant (0.05% Tween20) or 0.1% BSA to reduce background absorption.
- For release of antigens from biotinylated antibodies, glycine 0.1M pH 2.8 (low pH elution) is recommended. Heating above 70°C in reducing SDS-PAGE buffer also releases antigens, but biotinylated antibody and streptavidin as well.
- For coupling of nucleic acids, TE-buffer (pH 7.5) is recommended

IV. Product Usage

This product is stable for at least 1 year after purchasing date when stored at 2-8°C. Store in well closed vial and in upright position to prevent drying of the beads, this may result in a decrease of activity. Do not freeze the product! Vortex well before use. Wash the beads to remove preservatives that could interfere with your application. For washing, use the same volume as initially taken from the MagSi-STA vial or more.

V. Protocols

A) Bead preparation procedure

1. Resuspend beads by shaking/vortexing
2. Pipette the required volume of beads into a tube or microplate (10-20 μl is suitable as a starting point)
3. Collect beads by placing the tube or microplate on the magnet for 1-2 minutes
4. While tube/micro plate is still on the magnet, carefully remove supernatant without touching the bead pellet
5. Take tube/micro plate from the magnet and add washing buffer
6. Resuspend beads by vortexing or pipetting
7. Repeat step 3 – 5 at least 3 times
8. Finally resuspend the beads in a suitable buffer for your downstream, in a volume equal to the original bead volume.

B) General binding protocol

1. Add biotinylated molecule
2. Incubate for 30 minutes at room temperature
3. Collect beads by placing the tube or microplate on the magnet for 1-2 minutes
4. Wash beads 3-4 times with washing buffer
5. Resuspend the beads in a suitable buffer and volume for your downstream use.

C) Immunoprecipitation

1. Combine the antigen sample with 10 μg of biotinylated antibody. Dilute each sample to a minimum volume of 300 μl with cell lysis buffer or Binding/Wash Buffer. Incubate 1-2 hours at room temperature or overnight at 4°C with mixing.
2. Resuspend beads by shaking/vortexing
3. Add 25-50 μl of MagSi-STA beads into a 1.5 ml microcentrifuge tube.
4. Prepare the beads for binding by washing with binding buffer as described in a). Finally resuspend in 500 μl binding buffer.

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- Add the antigen sample/biotinylated antibody mixture to the 1.5 ml microcentrifuge tube containing pre-washed magnetic beads (3.) and incubate at RT for 30 minutes with mixing.
- Collect beads by placing the tube on the magnet for 1-2 minutes, pipette off and save the supernatant for analysis.
- Add 300 µl of binding/wash Buffer to the tube and gently mix. Collect the beads and then discard the supernatant. Repeat this step twice.
- Add 100 µl of elution buffer to the tube. For low pH elution, incubate the tube at room temperature with mixing for 5 minutes. For SDS-PAGE elution, add 100 µl of SDS-PAGE reducing sample buffer to the tube and heat the samples at 90°C for 10 minutes.
- Collect beads by placing the tube on the magnet for 1-2 minutes and transfer the supernatant containing target antigen.

*Low pH elution buffers are effective for most antibody-antigen interactions; however, to ensure efficient release of target antigen from the antibody, pre-rinse the beads with 300 µl 0.1% Tween-20 in water before adding low pH elution buffer.
 ** If SDS-PAGE buffer is selected for elution, the eluate will contain streptavidin monomers and dimers and biotinylated antibody along with target antigen.

D) Immunoassays

For direct capture, add MagSi-STA beads to a sample containing biotinylated antibodies. During a short incubation, the biotinylated molecules will bind to the beads. Collect the beads on a magnet and discard the supernatant. The beads are now ready to bind the antigen (analyte) from your sample.

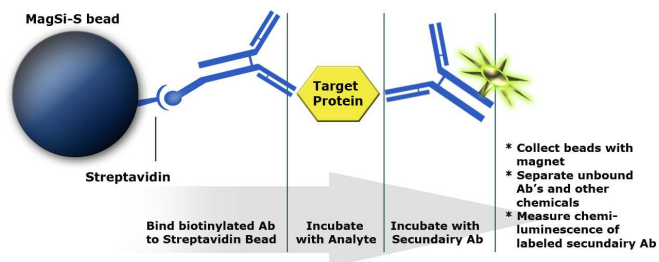


Fig.1: Principle of immunoassay based on MagSi-STA beads

For indirect capture, mix the biotinylated antibodies with your sample containing antigen before adding to the beads. Indirect capture can be

advantageous when binding conditions are slow or specific molecule orientation is needed.

Practical Notes:

- Optimize the quantity of beads for each application
- Use up to two-fold excess of the biotinylated molecule to saturate streptavidin on the beads

VI. Technical Data

Table 1: Specifications of MagSi-STA 600 and MagSi-STA 1.0

Product Name	MagSi-STA 600	MagSi-STA 1.0
Size	600 nm	1.0 µm
Concentration	10 mg/ml	
	beads/ml	
	8 - 20 · 10 ⁹	6 - 12 · 10 ⁹
Supplied product volume	2 ml, 10 ml, 100 ml	
Material	Magnetic silica beads with streptavidin covalently bound to the surface	
Size Distribution	D5-D95	
	500 - 900 nm	0.7 - 1.4 µm
Sedimentation		
Free Biotin binding capacity (pmol/mg)	3000 - 7000	
Solution additives	PBS (pH 7.4), 0.05% Tween20, 0.05% Sodium Azide (NaN ₃ , Toxic!)	
Storage	Store at 2-8°C	

VII. Additional Information

Disclaimer

For R&D use only. Not for drug, household or other uses. Product contains 0.05% sodium azide which is toxic. Avoid contact with the suspension buffer. When disposing the suspension buffer, flush with large amounts of water. Material Safety Data Sheet (MSDS) is available on our website at www.amsbio.com.

Order Information

Product	Volume	Art.No.
MagSi-STA 600	2 ml	MD16001
MagSi-STA 600	10 ml	MD18001
MagSi-STA 600	100 ml	MD19001
MagSi-STA 1.0	2 ml	MD01001
MagSi-STA 1.0	10 ml	MD03001
MagSi-STA 1.0	100 ml	MD04001

Related products

Product	Art.No.
MM-Separator M12 + 12	MD90001
MM-Separator M96	MD90002
MM-Separator 96 SBS	MD90005
MM-Separator 384 SBS	MD90006