COLO 205 Cell Line

Catalog No.: AMS.EP-CL-0066

Origin and General Characteristics

Cell Name	COLO 205 Cell Line
Organism	Homo sapiens, Human
Age	70 years
Tissue	Colon, derived from metastatic site: ascites
Morphology	Epithelial
Growth Properties	Mixed, Adherent And Suspension
Descriptions	The line was derived from tissue from the same patient as COLO 201. The cells are CSAp negative(CSAp-). The cells are positive for keratin by immunoperoxidase staining. COLO 205 cells express a 36000 dalton cell surface glycoprotein related to the GA733-2 tumor associated antigen.
Biosafety Level	1

Culture Conditions and Handling

Complete Growth Medium	RPMI-1640 +10% FBS +1% P/S
Subculturing	Remove and discard culture medium. These cells grow as a mixture of floating and adherent cells. Sometimes many cells are floating, they can be harvested by centrifugation of medium instead of discarding it. Add 1.0 to 2.0 mL of 0.25% (w/v) Trypsin-0.53mM EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 2 to 3 minutes). Add 4.0 to 6.0 mL of complete growth medium and aspirate cells by gently pipetting. Add appropriate aliquots of the cell suspension to new culture vessels. Incubate cultures at 37°C.
Subcultivation Ratio	1:2-1:4
Medium Renewal	Every 2 to 3 days
Cryopreservation	Freeze medium: 60% Basal medium+30% FBS+10% DMSO Storage temperature: Liquid nitrogen vapor phase
Culture Conditions	Atmosphere: Air, 95%; CO2, 5% Temperature: 37°C

Special Features of the Cell Line

Tumorigenic	Yes
Effects	Yes, in nude mice. Tumors developed within 21 days at 100% frequency(5/5) in nude mice inoculated subcutaneously with 10^7 cells.
Gene Expression	Carcinoembryonic Antigen(Cea) 1.5 To 4.1Ng/10^6 Cells/10 Days; Keratin; Interleukin 10(Il-10, Interleukin-10), The Cells Are Positive For Keratin By Immunoperoxidase Staining.
Applications	Transfection host.

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Recommendations for handling of cryopreserved cells

1. The cell is packaged by dry ice. When receiving the cell, please make sure that the vial is still frozen. If there is cell thawing in the tube, please take photo before experiment or storage.
2. If immediate culturing is not intended, the cryovial(s) must be stored in liquid nitrogen (-196°C) or at least at -80°C after arrival.

   If immediate culturing is intended, please follow these instructions:
3. Quickly thaw by rapid agitation in a 37°C water bath within 45-90 seconds. The water bath should have clean water containing an antimicrobial agent. As soon as the sample has thawed, remove the cryovial from the water bath.
   From now on, all operations should be carried out under aseptic conditions.
4. Transfer the cryovial to a sterile flow cabinet and wipe with 70% alcohol. Carefully open the vial and transfer the cell suspension into a 15 ml centrifuge tube containing 9 ml of cell complete medium (room temperature or 37°C).
5. In order to reduce cell damage, add 1ml of cell complete medium into cryovial, slightly pipette, then use a pipettor to add 1 ml of suspension into the centrifuge tube. Resuspend the cells carefully. Centrifuge at 300×g for 3 min and discard the supernatant. The centrifugation step may be omitted, but in this case the remains of the freeze medium have to be removed 24 hours later.
6. Resuspend the cells carefully in 10ml fresh cell culture medium and transfer them into one or two T25 cell culture flasks.

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