

# **Product Data Sheet**

# U118MG A-FLX™ FFPE Cell Pellet

## GENERAL INFORMATION

Product Name: U118MG A-FLX™ FFPE Cell Pellet

Reference Number: 3130-0110 Block

3130-0120 Slide (5µm)

3130-0130 FFPE scroll (20µm)

Date of Manufacturing: See product label
Lot Number: See product label
Intended Use: For research use only

**DESCRIPTION** 

Cell Line: U118MG
Tissue of Origin: Brain

Culturing Condition: DMEM supplemented with 10% FBS at 37°C with

5% CO<sub>2</sub>

Fixation Condition: 10% neutral buffered formalin (NBF) for 24 hours

at 24-27°C

Product Format:

Slide:

Block: Paraffin embedded block. Pellet thickness: ~2mm

One unstained section mounted on Superfrost™

Plus slide. Section thickness: 5µm

FFPE Scroll: One FFPE section in DNase/RNase free tube.

Section thickness: 20µm

#### **SCHEMATICS**

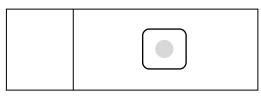


Illustration of an FFPE slide

#### STORAGE AND STABILITY

Storage Condition

Ambient temperature

2-8°C with desiccation (Recommended)

-20°C to -10°C

5 years

#### SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal productive equipment (PPE) when handling reagents and biological specimens.

### RECOMMENDED PROCEDURES

### Staining using FFPE slides:

- 1. Bake slides at 60°C for 30-60 min.
- 2. Deparaffinize two times in Xylene or Xylene substitute for 5 min each time
- 3. Rinse two times in 100% ethanol for 1 min each time.
- 4. Air dry slides or rehydrate in ethanol series (95% 2 min, 70% 2 min, 50% 2 min, 1XPBS 2 min).
- 5. Proceed to staining protocol.

### Biomolecule extraction from FFPE scrolls:

- 1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 10 sec.
- 2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 3. Repeat step 1 and 2.
- 4. Add 1ml 100% ethanol and mix by vortexing.
- 5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 6. Repeat step 4 and 5.
- 7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
- 8. Open the tube and dry at room temperature or up to 37°C for 10min, or until the ethanol has completely evaporated.
- 9. Proceed to extraction protocol.





\*Guideline for general applications, such as immunohistochemistry (IHC) or DNA in situ hybridization (ISH). Certain biomolecules may be less stable during storage.

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