

Product Data Sheet

BXPC3 A-FLX™ FFPE Cell Pellet

GENERAL INFORMATION

Product Name:	BXPC3 A-FLX™ FFPE Cell Pellet
Reference Number:	3120-0110 Block 3120-0120 Slide (5µm) 3120-0130 FFPE scroll (20µm)
Date of Manufacturing:	See product label
Lot Number:	See product label
Intended Use:	For research use only

DESCRIPTION

Cell Line:	BXPC3
Tissue of Origin:	Pancreas
Culturing Condition:	DMEM supplemented with 10% FBS at 37°C with 5% CO ₂
Fixation Condition:	10% neutral buffered formalin (NBF) for 24 hours at 24-27°C
Product Format:	
Block:	Paraffin embedded block. Pellet thickness: ~2mm
Slide:	One unstained section mounted on Superfrost™ Plus slide. Section thickness: 5µm
FFPE Scroll:	One FFPE section in DNase/RNase free tube. Section thickness: 20µm

SCHEMATICS

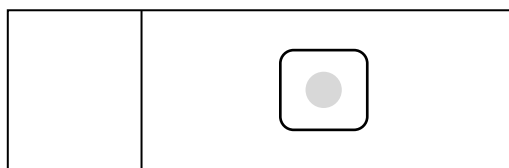


Illustration of an FFPE slide

STORAGE AND STABILITY

Storage Condition	Stability*
Ambient temperature	6 months
2-8 °C with desiccation (Recommended)	2 years
-20 °C to -10 °C	5 years

SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

RECOMMENDED PROCEDURES

Staining using FFPE slides:

1. Bake slides at 60°C for 30-60 min.
2. Deparaffinize two times in Xylene or Xylene substitute for 5 min each time.
3. Rinse two times in 100% ethanol for 1 min each time.
4. Air dry slides or rehydrate in ethanol series (95% 2 min, 70% 2 min, 50% 2 min, 1XPBS 2 min).
5. Proceed to staining protocol.

Biomolecule extraction from FFPE scrolls:

1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 10 sec.
2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
3. Repeat step 1 and 2.
4. Add 1ml 100% ethanol and mix by vortexing.
5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
6. Repeat step 4 and 5.
7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
8. Open the tube and dry at room temperature or up to 37°C for 10min, or until the ethanol has completely evaporated.
9. Proceed to extraction protocol.



*Guideline for general applications, such as immunohistochemistry (IHC) or DNA in situ hybridization (ISH). Certain biomolecules may be less stable during storage.

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