Product Data Sheet
A498 A-FLX™ FFPE Cell Pellet

GENERAL INFORMATION

Product Name:	A498 A-FLX™ FFPE Cell Pellet
Reference Number:	3100-0610 Block
	3100-0620 Slide (5µm)
	3100-0630 FFPE scroll (20µm)
Date of Manufacturing:	See product label
Lot Number:	See product label
Intended Use:	For research use only

DESCRIPTION

Cell Line:	A498
Tissue of Origin:	Kidney
Culturing Condition:	DMEM supplemented with 10% FBS at 37°C with 5% CO_2
Fixation Condition:	10% neutral buffered formalin (NBF) for 24 hours at 24-27°C
Product Format:	
Block:	Paraffin embedded block. Pellet thickness: ~2mm
Slide:	One unstained section mounted on Superfrost™ Plus slide. Section thickness: 5µm
FFPE Scroll:	One FFPE section in DNase/RNase free tube. Section thickness: 20µm

SCHEMATICS

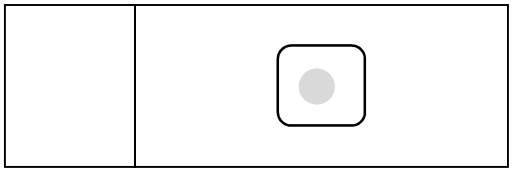

Illustration of an FFPE slide

STORAGE CONDITION

2-8 ˚C with desiccation (Recommended)

*STABILITY

Block	3 years
Scroll	1 year
Slide	1 year

*Guideline for general applications, such as immunohistochemistry (IHC) or DNA in situ hybridization (ISH). Certain biomolecules may be less stable during storage.

SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal productive equipment (PPE) when handling reagents and biological specimens.

RECOMMENDED PROCEDURES

Staining using FFPE slides:

1. Bake slides at 60°C for 30-60 min.
2. Deparaffinize two times in Xylene or Xylene substitute for 5 min each time.
3. Rinse two times in 100% ethanol for 1 min each time.
4. Air dry slides or rehydrate in ethanol series (95% 2 min, 70% 2 min, 50% 2 min, 1XPBS 2 min).
5. Proceed to staining protocol.

Biomolecule extraction from FFPE scrolls:

1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 10 sec.
2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
3. Repeat step 1 and 2.
4. Add 1ml 100% ethanol and mix by vortexing.
5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
6. Repeat step 4 and 5.
7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
8. Open the tube and dry at room temperature or up to 37°C for 10min, or until the ethanol has completely evaporated.
9. Proceed to extraction protocol.

AMSBIO | www.amsbio.com | info@amsbio.com

UK & Rest of the World
184 Park Drive, Milton Park
Abingdon, UK
T: +44 (0)1235 828 200
F: +44 (0) 1235 820 482

North America
1035 Cambridge Street,
Cambridge, MA 02141
T: +1 (617) 945-5033 or
T: +1 (800) 987-0985
F: +1 (617) 945-8218

Germany
Bockenheimer Landstr. 17/19
60325 Frankfurt/Main
T: +49 (0) 69 779099
F: +49 (0) 69 13376880

Switzerland
Centro Nord-Sud 2E
CH-6934 Bioggio-Lugano
T: +41(0) 91 604 55 22
F: +41(0) 91 605 17 85