

786-O FFPE Cell Pellet

GENERAL INFORMATION

Product Name:	786-O FFPE Cell Pellet
Reference Number:	3100-0310 Block 3100-0320 Slide (5µm) 3100-0330 FFPE scroll (20µm)
Date of Manufacturing:	See product label
Lot Number:	See product label
Intended Use:	For research use only

DESCRIPTION

Cell Line:	786-O
Tissue of Origin:	Kidney
Culturing Condition:	DMEM supplemented with 10% FBS at 37°C with 5% CO ₂
Fixation Condition:	10% neutral buffered formalin (NBF) for 24 hours at 24-27°C
Product Format:	
Block:	Paraffin embedded block. Pellet thickness: ~2mm
Slide:	One unstained section mounted on Superfrost™ Plus slide. Section thickness: 5µm
FFPE Scroll:	One FFPE section in DNase/RNase free tube. Section thickness: 20µm

SCHEMATICS

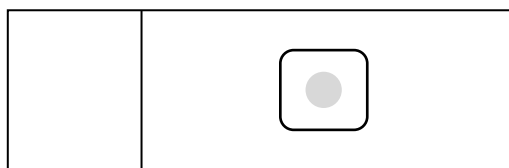


Illustration of an FFPE slide

STORAGE AND STABILITY

Storage Condition	Stability*
Ambient temperature	6 months
2-8 °C with desiccation (Recommended)	2 years
-20 °C to -10 °C	5 years

*Guideline for general applications, such as immunohistochemistry (IHC) or DNA in situ hybridization (ISH). Certain biomolecules may be less stable during storage.

SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

RECOMMENDED PROCEDURES

Staining using FFPE slides:

1. Bake slides at 60°C for 30-60 min.
2. Deparaffinize two times in Xylene or Xylene substitute for 5 min each time.
3. Rinse two times in 100% ethanol for 1 min each time.
4. Air dry slides or rehydrate in ethanol series (95% 2 min, 70% 2 min, 50% 2 min, 1XPBS 2 min).
5. Proceed to staining protocol.

Biomolecule extraction from FFPE scrolls:

1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 10 sec.
2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
3. Repeat step 1 and 2.
4. Add 1ml 100% ethanol and mix by vortexing.
5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
6. Repeat step 4 and 5.
7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
8. Open the tube and dry at room temperature or up to 37°C for 10min, or until the ethanol has completely evaporated.
9. Proceed to extraction protocol.



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