

## HL60 FFPE Cell Pellet

### GENERAL INFORMATION

|                        |  |
|------------------------|--|
| Product Name:          | HL60 FFPE Cell Pellet  |
| Reference Number:      | 3070-0410 Block<br>3070-0420 Slide (5µm)<br>3070-0430 FFPE scroll (20µm) |
| Date of Manufacturing: | See product label  |
| Lot Number:            | See product label  |
| Intended Use:          | For research use only  |

### DESCRIPTION

|                      |   |
|----------------------|---|
| Cell Line:           | HL60  |
| Tissue of Origin:    | Leukemia  |
| Culturing Condition: | Multiple  |
| Fixation Condition:  | 10% neutral buffered formalin (NBF) for 24 hours at 24-27°C                     |
| Product Format:      |   |
| Block:               | Paraffin embedded block. Pellet thickness: ~2mm                                 |
| Slide:               | One unstained section mounted on Superfrost™ Plus slide. Section thickness: 5µm |
| FFPE Scroll:         | One FFPE section in DNase/RNase free tube. Section thickness: 20µm              |

### SCHEMATICS

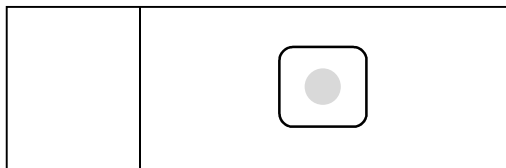


Illustration of an FFPE slide

\*Guideline for general applications, such as immunohistochemistry (IHC) or DNA in situ hybridization (ISH). Certain biomolecules may be less stable during storage

### SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

### RECOMMENDED PROCEDURES

#### Staining using FFPE slides:

1. Bake slides at 60°C for 30-60 min.
2. Deparaffinize two times in Xylene or Xylene substitute for 5 min each time.
3. Rinse two times in 100% ethanol for 1 min each time.
4. Air dry slides or rehydrate in ethanol series (95% 2 min, 70% 2 min, 50% 2 min, 1XPBS 2 min).
5. Proceed to staining protocol.

#### Biomolecule extraction from FFPE scrolls:

1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 10 sec.
2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
3. Repeat step 1 and 2.
4. Add 1ml 100% ethanol and mix by vortexing.
5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
6. Repeat step 4 and 5.
7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
8. Open the tube and dry at room temperature or up to 37°C for 10min, or until the ethanol has completely evaporated.
9. Proceed to extraction protocol.

### STORAGE AND STABILITY

|                   |                                       |
|-------------------|---------------------------------------|
| Storage Condition | 2-8 °C with desiccation (Recommended) |
|-------------------|---------------------------------------|

### STABILITY

|        |         |
|--------|---------|
| Block  | 3 years |
| Scroll | 1 year  |
| Slide  | 1 year  |

AMSBIO | [www.amsbio.com](http://www.amsbio.com) | [info@amsbio.com](mailto:info@amsbio.com)