

HT144 FFPE Cell Pellet

GENERAL INFORMATION

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|------------------------|---|
| Product Name: | HT144 FFPE Cell Pellet |
| Reference Number: | 3060-0710 Block 3060-0720 Slide (5µm) 3060-0730 Scroll (20µm) |
| Date of Manufacturing: | See product label |
| Lot Number: | See product label |
| Intended Use: | For research use only |

DESCRIPTION

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|----------------------|---|
| Cell Line: | HT144 |
| Tissue of Origin: | Skin |
| Culturing Condition: | DMEM supplemented with 10% FBS at 37°C with 5% CO ₂ |
| Fixation Condition: | 10% neutral buffered formalin (NBF) for 24 hours at 24-27°C |
| Product Format: | |
| Block: | Paraffin embedded block. Pellet thickness: ~2mm |
| Slide: | One unstained section mounted on Superfrost™ Plus slide. Section thickness: 5µm |
| FFPE Scroll: | One FFPE section in DNase/RNase free tube. Section thickness: 20µm |

SCHEMATICS

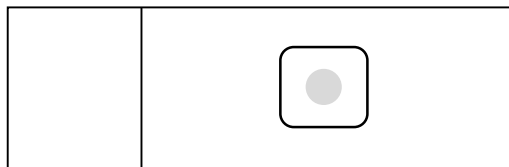


Illustration of an FFPE slide

STORAGE CONDITION

2-8 °C with desiccation (Recommended)

*STABILITY

| | |
|--------|---------|
| Block | 5 years |
| Scroll | 1 year |
| Slide | 1 year |

SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

RECOMMENDED PROCEDURES

Staining using FFPE slides:

1. Bake slides at 60°C for 30-60 min.
2. Deparaffinize two times in Xylene or Xylene substitute for 5 min each time.
3. Rinse two times in 100% ethanol for 1 min each time.
4. Air dry slides or rehydrate in ethanol series (95% 2 min, 70% 2 min, 50% 2 min, 1XPBS 2 min).
5. Proceed to staining protocol.

Biomolecule extraction from FFPE scrolls:

1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 10 sec.
2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
3. Repeat step 1 and 2.
4. Add 1ml 100% ethanol and mix by vortexing.
5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
6. Repeat step 4 and 5.
7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
8. Open the tube and dry at room temperature or up to 37°C for 10min, or until the ethanol has completely evaporated.
9. Proceed to extraction protocol.



*Guideline for general applications, such as immunohistochemistry (IHC) or DNA in situ hybridization (ISH). Certain biomolecules may be less stable during storage.