amsbio

# H1650 FFPE Cell Pellet

### GENERAL INFORMATION

Product Name:	H1650 FFPE Cell Pellet	
Reference Number:	3020-1210 Block	
	3020-1220 Slide (5µm)	
	3020-1230 FFPE scroll (20µm)	
Date of Manufacturing:	See product label	
Lot Number:	See product label	
Intended Use:	For research use only	

### DESCRIPTION

Cell Line:	H1650
Tissue of Origin:	Lung
Culturing Condition:	RPMI-1640 supplemented with 10% FBS at 37°C with 5% $\text{CO}_2$
Fixation Condition:	10% neutral buffered formalin (NBF) for 24 hours at 24-27°C
Product Format:	
Block:	Paraffin embedded block. Pellet thickness: ~2mm
Slide:	One unstained section mounted on Superfrost™ Plus slide. Section thickness: 5µm
FFPE Scroll:	One FFPE section in DNase/RNase free tube. Section thickness: 20µm

### SCHEMATICS

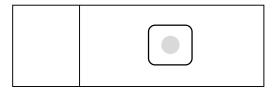


Illustration of an FFPE slide

### STORAGE CONDITION

2-8 °C with desiccation (Recommended)

### \*STABILITY

Block	5 y
Scroll	1 y
Slide	1 y

## 5 years 1 year 1 year

**Product Data Sheet** 

### SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal productive equipment (PPE) when handling reagents and biological specimens.

### **RECOMMENDED PROCEDURES**

### Staining using FFPE slides:

- 1. Bake slides at 60°C for 30-60 min.
- 2. Deparaffinize two times in Xylene or Xylene substitute for 5 min each time.
- 3. Rinse two times in 100% ethanol for 1 min each time.
- 4. Air dry slides or rehydrate in ethanol series (95% 2 min, 70% 2 min, 50% 2 min, 1XPBS 2 min).
- 5. Proceed to staining protocol.

### Biomolecule extraction from FFPE scrolls:

- 1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 10 sec.
- 2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 3. Repeat step 1 and 2.
- 4. Add 1ml 100% ethanol and mix by vortexing.
- 5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 6. Repeat step 4 and 5.
- 7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
- 8. Open the tube and dry at room temperature or up to 37°C for 10min, or until the ethanol has completely evaporated.
- 9. Proceed to extraction protocol.

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\*Guideline for general applications, such as immunohistochemistry (IHC) or DNA in situ hybridization (ISH). Certain biomolecules may be less stable during storage.

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